**EFFECT OF DIETARY SUPPLEMENTS ON BONE HEALING IN GRAFTED DEFECTS: AN EXPERIMENTAL STUDY IN RATS**

**ABSTRACT**

**Objective:** The aim of this investigation was to evaluate the potential effects of the systemically delivered combination of calcium, zinc and vit-d supplementation of the locally applied alloplastic bone graft on the surgically created tibial defects in rat model.

**Material and Methods:** 28 male Wistar albino rats were used in this study. In each animal, bone defects were created in the tibias. The animals were divided into four groups. In Group 1 (Control Group) rats were fed with standard rat diet. In Group 2 (Calcium) rats received calcium carbonate (15 mg/kg) suspended in saline. In Group 3 (Calcium/Zinc) rats received calcium carbonate (15 mg/kg) and zinc sulfate (4 mg/kg) suspended in saline. In Group 4 (Calcium/Vitamin D) rats received calcium carbonate (15 mg/kg) and Vitamin D (500 IU/kg) suspended in olive oil.

**Results:** The animals were sacrificed on the 21st postoperative day. Histopathological analysis of

samples was performed to evaluate the process of osteoblastic activity(OA), matrix formation(MF), trabecular bone formation(TBF) and myeloid tissue(MT) in bone defects. Total amounts of OA, MF, TBF and MT in Group 2 (p = 0,002), Group 3 (p = 0,002), and Group 4 (p = 0,001) were significantly higher than in Group 1. The total amounts of Group 4 were significantly different than Group 1 and Group 2. No statistically differences were observed in total amounts of OA, MF, TBF and MT between Group 3 and Group 4.

**Conclusion:** The results of the present study indicated that the oral calcium carbonate supplementation combination with Zinc may have systemic effects on accelerating bone regeneration in alloplastic bone grafted tibial defects, but the Group 4 was also effective with no statiscally differences between Group 3 and Group 4. Further human studies involving long-term follow up and different type of bone grafts should be conducted.

**INTRODUCTION**

Bone is composed of two main structural types: primary bone and lamellar (secondary) bone. Bone repair process may take months or years . The recovery of hard tissues lost during the treatment of pathological processes and traumatic lesions has been extensively studied, and different approaches have been suggested (1,2). Autogenous bone graft is considered to be the gold standard for replacement of lost tissue (3). Advancements in surgical techniques to collect human bone for autogenous grafting are not able to keep pace with the evolution in the production of synthetic materials, such as calcium phosphate cements which have been successfully used for bone repair in the last decade (4).

Bone healing involves complex processes of cell and tissue proliferation and differentiation. Many players are involved, including growth factors, inflammatory cytokines, antioxidants, bone breakdown (osteoclast) and bone‐building (osteoblast) cells, hormones, amino acids, and uncounted nutrients (5).

Calcium (Ca) is an essential nutrient for normal growth and development. Adequate dietary calcium builds the skeleton and helps to prevent skeletal disorders during childhood and adolescence (6). The main bone minerals are calcium and phosphorus, in the form of calcium hydroxyapatite (HA) crystals. This HA compound plays an important role in regulating the elastic stiffness and tensile strength of bone (7). The building and rebuilding of bone tissue requires adequate supplies of both calcium and phosphorus, which can be supplied from diet and bone reserves. Inadequate dietary calcium during the critical growth and building period may result in failure to reach peak bone mass (8).

Vitamin-D (Vit.D) is the primary regulator of calcium absorption and without adequate vit-D calcium blood level drops making less calcium available for fracture healing. Further, we now know that vitamin D, in conjunction with vitamin K, stimulates the transformation of fracture site stem cells to bone building osteoblasts (9).

Zinc has an important role in osteogenesis. Bone has one of the highest concentrations of zinc of all tissues. Several zinc-dependent enzymes and hormones are involved in bone metabolism. One of them is stimulation of the activity of alkaline phosphatase (ALP), which is involved in bone mineral deposition. The cellular mechanism of zinc action has been demonstrated to stimulate proliferation and differentiation in osteoblastic cells and to inhibit formation of osteoclastic cells. It causes the decrease of cellular zinc content and protein synthesis in bone tissues (10).

The aim of this study was to investigate the effects of dietary calcium with zinc and vit.D supplements in the healing of calcium phosphate bone grafted tibial defects in rat model.

**MATERIALS AND METHODS**

This research was conducted with the approval of the Dicle University Animal Care and Use Ethical Committee (permit no: 2011-31) and rats were purchased from the Dicle University Center of Experimental Medical Research. 28 male Wistar albino rats weighting 200-215 gr were used in this study. Rats were housed in standard cages in a temperature (22 ± 2°C) and humidity (55 ± 5%) controlled room that maintained with a 12 h/12 h light/dark cycle throughout the experiment. The animals consumed a commercial standard laboratory diet and tap water *ad libitum*. The animals were anesthetized by intraperitoneal injection 5 mg/kg of Ketamine (Ketalar®, Eczacıbaşı, Turkey) and Xylazine HCL (Rompun®, Bayer, Turkey). The skin was shaved and scrubbed with an antiseptic solution (1% iodine). A 2 cm longitudinal incision was made along the frontal aspect of tibia and flaps were raised to expose the bone tissue. 10 mm long, 3 mm deep and 2 mm wide circular standard bone defects involving cortical and cancellous bone layers were performed by round dental bur with low rpm, under irrigation sterile saline and suction. The defects were filled with the alloplastic bone graft material (Beta Tricalcium Phosphate (β – TCP) Suprabone®, 1-2 mm particle size, BMT Calsis, Turkey). The periosteum was closed using silk 3-0 sutures (Ethicon®, Edinburgh, UK). Rats were divided into four experimental groups ( Table 1). Calcium Carbonate (CaCO3) (Calcium Sandoz Forte®, Novartis, Turkey), Zinc Sulfate (Zinco® 15mg, Berko, Turkey) and Vitamin D (Devit-3, Deva, Turkey) were administered intragastrically one time per day during the experimental period (Fig1).

**Group 1:** Rats were fed with standard rat diet.

**Group 2:** Rats received CaCO3 (15 mg/kg) suspended in saline.

**Group 3:** Rats received CaCO3 (15 mg/kg) and Zinc Sulfate (4 mg/kg) suspended in saline.

**Group 4:** Rats received CaCO3 (15 mg/kg) and Vitamin D (500 IU/kg) suspended in olive oil.

The rats were sacrified by an overdose of sodium thiopental (Pentothal®) to assess the bone healing response on the 21st postoperative day. A skin incision with a periosteal flap was used to expose the tibial bone and the previously grafted site was excised widely. The samples were fixed in 10% buffered neutral formalin for 24 hours and then decalcified in a formic acid-hydrochloride acid combination for 24 hours. After rinsing with tap water, the samples were dehydrated in increasing concentrations of ethanol and embedded in paraffin. Tissue sections of 5–7 *μ*m thickness were prepared in the transverse plane and stained using Hematoxylin-eosin and Masson’s trichrome staining methods. The histopathological scoring system (11) was used for evaluation of bone regeneration process to pattern on the grade of osteoblastic activity, matrix formation, trabecular bone formation and myeloid tissue (Table 2,3). The data were analyzed statistically using the Statistical Package for the Social Sciences, ver. 12.0 (SPSS, Chicago, IL, USA). The results are expressed as means ± standard deviation (SD). Differences were evaluated using the Mann–Whitney U-test, and *p* values < 0.05 were considered to indicate statistical significance (Table 4).

**RESULTS AND DISCUSSION**

**Group 1 (Standard Diet) (Control):** Closure of the defect areas were seen acceleration of the stem cell formation could be seen by increasing of the myeloid tissue and reticular fiber activity and also increased new lamellar bone formation was observed according to maturation of the bone matrix by osteoblasts. Increased osteoblastic activity was found between spongeous and trabecular bone tissues (Fig 2A).

**Group 2 (Ca):** Low osteoblastic activity was evidenced by poor osteoblasts and osteosits lining periferarly of the trabecular bone. Additionally myeloid tissue formation in bone narrow was higher more than matrix and trabecular bone formation. (Fig 2B).

**Group 3 (Ca+Zinc):** Despite the presence of new trabecular bone formation and matrix formation, bone healing was not significant. Osteoblasts were lining as cubical between spongeous and trabecular bone tissues. Nevertheless increased lipid cell formation and myeloid tissue formation were observered with increased mitotic activity in the myeloid tissue. There were also seen some small vessels in the trabecular bone with increased osteocytic activity (Fig 2C).

**Group 4 (Ca+Vit.D):** The samples in this group showed increased bone lamellae formation in the trabecular bone and increased osteoblastic activity in the lacunar area. Matrix formation and osteoblastic activity were seen strongly significant in the osteoid region (Fig 2D).

Total amounts of Osteoblastic Activity, Matrix Formation, Trabecular Bone Formation and Myeloid Tissue in Group 2 (p = 0,002), Group 3 (p = 0,002), and Group 4 (p = 0,001) were significantly higher than in Group 1 (Control Group). The values of Group 3 and Group 4 were higher than Group 2 but no statiscally significant. The total amounts of Group 3 were significantly different than Group 1 and Group 2. No statistically differences were observed in total amounts of Osteoblastic Activity, Matrix Formation, Trabecular Bone Formation and Myeloid Tissue between Group 3 and Group 4. The Trabecular Bone Formation amounts in Group 4 were found less than in Groups 2 and 3 and it was statiscally significant (p = 0,01), but Osteoblastic Activity values were not significantly different. The distributions of the histological scores are shown in Figure 3.

Calcium supplements are becoming an important source of dietary calcium and a basic defense against osteopenia. Oral calcium supplement is a very popular method of maintenance inadequate dietary calcium because of its cheap cost, convenient way of intake, and the minimal side effects. Generally, calcium supplementation is recognized as accelerating bone formation with less inhibition of bone resorption (12). The role of calcium-regulating hormones, vitamin D, parathyroid hormone and calcitonin, and ions such as calcium and phosphate in mediating osteoinduction is not known. There is evidence that vitamin D deficiency results in abnormalities in skeletal growth, remodeling, and mineralization. Human studies, in fact, suggest that for best fracture healing both calcium and vitamin D should be obtained in optimum daily levels. Most of us consume plenty of phosphorus and often too much if the diet is high in processed foods and colas. However, the elderly, dieters, and those on low protein diets often do not consume enough phosphorus to meet the needs of new bone formation (13). Thus we evaluated in this study the efficacy of dietary Calcium supplementation with/without Zinc and Vit-D in healing of the alloplastic bone grafted rat tibial defects.

Hong et al. evaluated the effects of the oral calcium and cholecalciferol combination (Vit.D/Ca) supplementation and of the locally applied biphasic calcium alloplast on alveolar sockets at the early healing stage in a dog model. The Vit.D/Ca-treated subjects revealed significantly more new bone formation (P < 0.05), higher bone density (P < 0.05) and significantly less vertical ridge reduction (P < 0.05) in the healing sockets than those without Vit.D/Ca treatment (14). At an early stage of the defect region, results of this study demonstrate that taking calcium supplementation significantly stimulates bone regeneration and better new bone formation than non-supplemented group. The information supports the idea that systemic application of Vit.D/Ca may encourage an early bone healing in investigational areas for the tested animals. Shuid et al. carried out in a study on the effects of calcium supplementation on the late phase healing of fractured osteoporotic bone using an ovariectomized rat model. They concluded that calcium supplements may appear to improve fracture healing of osteoporotic bone but failed to improve strength (15). Qin et al. evaluated in their study the effect of the mixture of calcium carbonate and calcium citrate, on the mandibular alveolar bone debilitation of pubertal rats. They concluded that the mixture of calcium carbonate and calcium citrate had a positive effect on bone debilitation to a certain extent in growth-period rats (16).Turner et al. implanted demineralized allogeneic bone matrix (DABM) from vitamin D-deficient rats (-D rats) into normal rats (+D rats). Bone formation (P < 0,01) and total implant mineralization (P < 0.001) were significantly reduced in implants from -D rats, and the reductions corresponded with a decline in the number of osteoblasts (P < 0.05)(17).

In the present study between the Calcium Group (15mg/kg CaCO3) and Ca/Vit.D Group (15mg/kg CaCO3 and 500 IU/kg Vit.D) were not seen significanlty differences in bone healing. It shows that sparingly Vit.D is enough for intestinal calcium absorption. Calhoun et al. indicated that dietary zinc deficiency caused a retardation of ectopic bone formation and a significant reduction of in situ zinc and calcium concentration. Dietary zinc repletion to zinc-deficient animals restored the zinc concentration in ectopic bone to a level comparable to that of zinc-sufficient animals (18).Hosea et al. investigated the effects of dietary zinc deficiency and diet restriction on bone development in growing rats. Rats were fed either a zinc-deficient diet ad libitum (<1 mg zinc/kg) or nutritionally complete diet (30 mg zinc/kg) either ad libitum (CTL) or pair-fed to the intake of the ZD group (DR; diet-restricted) for 3 weeks (deficiency phase) and then all groups were fed the zinc adequate diet ad libitum for 3, 7, or 23 days (repletion phase). Zinc deficiency was associated with a greater impairment in bone development than diet restriction, and both deficiencies limited bone recovery during repletion in growing rats(19).

According to our observation we can say systemically administration of Zinc or Vit.D with Ca increased the new bone regeneration in grafted rat tibial defects more than Ca alone supplementation but only Zinc/Ca group was statiscally different (p= 0.005). Igarashi et al. investigated the effect of zinc acexamate (10.0 mg Zn/100 g) for 28 days on fracture healing of the femoral-diaphyseal tissues in rats. Calcium content and alkaline phosphatase activity in the femoral-diaphyseal tissues were significantly decreased in rats with fracture healing. Femoral mineral density in rats with fracture healing was significantly increased by the administration of zinc acexamate. Femoral-diaphyseal zinc content was significantly decreased in rats with fracture healing (20). Jones et al. evaluated whether alimentary zinc supplementation and depletion affect bone healing of calvarial defects implanted with osteopromotive substances in adult rats.Experimentally created bone defects were filled with demineralized bone matrix (DBM), autogenous bone chips, or were left as unfilled controls The rats were divided into three groups of 20 rats each and received a semi-synthetic diet containing 20, 60, or 120 mg zinc/kg. Defects filled with DBM were significantly stronger and exhibited significantly more new bone formation than defects filled with autogenous bone or unfilled controls(21). In our study the rats in Group Ca/Zinc were fed with 4 mg/kg Zinc (Zinc Sulfate (15mg Zn/100 ml) for 21 days. The alloplastic bone grafted tibial tissues of Ca/Zinc Group revealed significantly more new bone formation compared than those Control Group and Ca Group.

In conclusion it has been demonstrated that the oral administration of calcium carbonate is effective in new bone process and Zinc sulfate has a sinergic stimulatory effect with calcium carbonate on healing of experimental created and alloplastic bone grafted tibial defects in rats.

**REFERENCES**

1. Burkitt HG, Young B, Heath JW. Wheater Histologia Funcional**.** 3rd edition. Rio de Janeiro: Guanabara Koogan; 1994.
2. Schilling T, Müller M, Minne HW, Ziegler R. Influence of inflammationmediated osteopenia on the regional acceleratory phenomenon and the systemic acceleratory phenomenon during healing of a bone defect in the rat**.** Calcif Tissue Int 63:160-166, 1998.
3. Puricelli E, Chem RC: Reconstrução de mandíbula com fíbula**.** Rev Gaúcha Odontol 33:124-127, 1985.
4. Kurashina K, Kurita H, Kotani A, Kobayashi S, Kyoshima K, Hirano M. Experimental cranioplasty and skeletal augmentation using an alphatricalcium phosphate/dicalcium phosphate dibasic/tetracalcium phosphate monoxide cement: a preliminary short-term experiment in rabbits**.** Biomaterials, 19:701-706, 1998.
5. Marsh DR, Li G. The biology of fracture healing: Optimising outcome, British Medical Bulletin, 55(4):856-869, 1999.
6. Matkovic V. Calcium metabolism and calcium requirement during skeletal modeling and consolidation of bone mass. Am J Clin Nutr 54:245-260, 1991.
7. Simşek A, Senköylü A, Cila E, Uğurlu M, Bayar A, Oztürk AM, Işikli S, Muşdal Y, and Yetkin H. Is there a correlation between severity of trauma and serum trace element levels?, Acta Orthop Traumatol Turc 40(2):140-143, 2006.
8. Key JA, Odell RT. Failure of excess minerals in diet to accelerate the healing of experimental fractures. J Bone Joint Surg 37-A(1):37-44, 1955.
9. Steier A, Gedalia I, Schwarz A, and Rodan A. Effect of vitamin D2 and fluoride on experimental bone fracture healing in rats. J Dent Res 46(4):675-680, 1967.
10. Yamaguchi M, Yamaguchi R. Action of zinc on bone metabolism in rats. Biochem Pharmacol 35:773–777, 1986.
11. Yeh WL, Lin SS, Yuan LJ, Lee KF, Lee MY, Ueng SW.Effects of hyperbaric oxygen treatment on tendon graft and tendon-bone integration in bone tunnel: biochemical and histological analysis in rabbits. J Orthop Res 2007;25:636-45
12. Recker RR. Prevention of osteoporosis: calcium nutrition. Osteoporosis Int 3(1):163-5, 1993.
13. Heaney RP and Nordin BEC. Calcium effects on phosphorus absorption: Implications for the prevention and co-therapy of osteoporosis. J Am Coll Nutr 21(3):239-244, 2002.
14. Hong H-H, Chou T-A, Yang J-C, Chang C-J. The potential effects of cholecalciferol on bone regeneration in dogs. Clin. Oral Impl. Res 23: 1187–1192, 2012.
15. [Shuid AN](http://www.ncbi.nlm.nih.gov/pubmed?term=Shuid%20AN%5BAuthor%5D&cauthor=true&cauthor_uid=20572125), [Mohamad S](http://www.ncbi.nlm.nih.gov/pubmed?term=Mohamad%20S%5BAuthor%5D&cauthor=true&cauthor_uid=20572125), [Mohamed N](http://www.ncbi.nlm.nih.gov/pubmed?term=Mohamed%20N%5BAuthor%5D&cauthor=true&cauthor_uid=20572125), [Fadzilah FM](http://www.ncbi.nlm.nih.gov/pubmed?term=Fadzilah%20FM%5BAuthor%5D&cauthor=true&cauthor_uid=20572125), [Mokhtar SA](http://www.ncbi.nlm.nih.gov/pubmed?term=Mokhtar%20SA%5BAuthor%5D&cauthor=true&cauthor_uid=20572125), [Abdullah S](http://www.ncbi.nlm.nih.gov/pubmed?term=Abdullah%20S%5BAuthor%5D&cauthor=true&cauthor_uid=20572125), [Othman F](http://www.ncbi.nlm.nih.gov/pubmed?term=Othman%20F%5BAuthor%5D&cauthor=true&cauthor_uid=20572125), [Suhaimi F](http://www.ncbi.nlm.nih.gov/pubmed?term=Suhaimi%20F%5BAuthor%5D&cauthor=true&cauthor_uid=20572125), [Muhammad N](http://www.ncbi.nlm.nih.gov/pubmed?term=Muhammad%20N%5BAuthor%5D&cauthor=true&cauthor_uid=20572125), [Soelaiman IN](http://www.ncbi.nlm.nih.gov/pubmed?term=Soelaiman%20IN%5BAuthor%5D&cauthor=true&cauthor_uid=20572125). Effects of Calcium Supplements on Fracture Healing in a Rat Osteoporotic Model. J. Orthop. Res 28: 1651–1656, 2010.
16. [Man Qin](http://link.springer.com/search?facet-author=%22Man+Qin%22), [Zuyan Zhang](http://link.springer.com/search?facet-author=%22Zuyan+Zhang%22), [Kenshi Maki](http://link.springer.com/search?facet-author=%22Kenshi+Maki%22), [Mariko Naito](http://link.springer.com/search?facet-author=%22Mariko+Naito%22), [Akiko Morimoto](http://link.springer.com/search?facet-author=%22Akiko+Morimoto%22), [Mitsutaka Kimura](http://link.springer.com/search?facet-author=%22Mitsutaka+Kimura%22). The effect of calcium supplement given with a mixture of calcium carbonate and calcium citrate on the mandibular alveolar bone of pubertal rats. [Journal of Bone and Mineral Metabolism](http://link.springer.com/journal/774) 16(2):88-95, 1998.
17. [Turner RT](http://www.ncbi.nlm.nih.gov/pubmed?term=Turner%20RT%5BAuthor%5D&cauthor=true&cauthor_uid=3260604), [Farley J](http://www.ncbi.nlm.nih.gov/pubmed?term=Farley%20J%5BAuthor%5D&cauthor=true&cauthor_uid=3260604), [Vandersteenhoven JJ](http://www.ncbi.nlm.nih.gov/pubmed?term=Vandersteenhoven%20JJ%5BAuthor%5D&cauthor=true&cauthor_uid=3260604), [Epstein S](http://www.ncbi.nlm.nih.gov/pubmed?term=Epstein%20S%5BAuthor%5D&cauthor=true&cauthor_uid=3260604), [Bell NH](http://www.ncbi.nlm.nih.gov/pubmed?term=Bell%20NH%5BAuthor%5D&cauthor=true&cauthor_uid=3260604), [Baylink DJ](http://www.ncbi.nlm.nih.gov/pubmed?term=Baylink%20DJ%5BAuthor%5D&cauthor=true&cauthor_uid=3260604). Demonstration of Reduced Mitogenic and Osteoinductive Activities in Demineralized Allogeneic Bone Matrix from Vitamin D-deficient Rats [J Clin Invest](http://www.ncbi.nlm.nih.gov/pubmed/?term=Demonstration+of+Reduced+Mitogenic+and+Osteoinductive+Activities+in+Demineralized+Allogeneic+Bone+Matrix+from+Vitamin+D-deficient+Rats)  82(1):212-7, 1988.
18. [Calhoun NR](http://www.ncbi.nlm.nih.gov/pubmed?term=Calhoun%20NR%5BAuthor%5D&cauthor=true&cauthor_uid=1056573), [Smith JC Jr](http://www.ncbi.nlm.nih.gov/pubmed?term=Smith%20JC%20Jr%5BAuthor%5D&cauthor=true&cauthor_uid=1056573), [Becker KL](http://www.ncbi.nlm.nih.gov/pubmed?term=Becker%20KL%5BAuthor%5D&cauthor=true&cauthor_uid=1056573). The effect of zinc ectopic bone formation. [Oral Surg Oral Med Oral Pathol](http://www.ncbi.nlm.nih.gov/pubmed/1056573) 39(5):698-706, 1975.
19. [Hosea HJ](http://www.ncbi.nlm.nih.gov/pubmed?term=Hosea%20HJ%5BAuthor%5D&cauthor=true&cauthor_uid=15044713), [Taylor CG](http://www.ncbi.nlm.nih.gov/pubmed?term=Taylor%20CG%5BAuthor%5D&cauthor=true&cauthor_uid=15044713), [Wood T](http://www.ncbi.nlm.nih.gov/pubmed?term=Wood%20T%5BAuthor%5D&cauthor=true&cauthor_uid=15044713), [Mollard R](http://www.ncbi.nlm.nih.gov/pubmed?term=Mollard%20R%5BAuthor%5D&cauthor=true&cauthor_uid=15044713), [Weiler HA](http://www.ncbi.nlm.nih.gov/pubmed?term=Weiler%20HA%5BAuthor%5D&cauthor=true&cauthor_uid=15044713). Zinc-deficient have more limited bone recovery during repletion than diet-restricted rats. [Exp Biol Med (Maywood)](http://www.ncbi.nlm.nih.gov/pubmed/?term=Zinc-Deficient+Rats+Have+More+Limited+Bone+Recovery+During+Repletion+Than+Diet-Restricted+Rats) 229(4):303-11, 2004.
20. [Igarashi A](http://www.ncbi.nlm.nih.gov/pubmed?term=Igarashi%20A%5BAuthor%5D&cauthor=true&cauthor_uid=10323487), [Yamaguchi M](http://www.ncbi.nlm.nih.gov/pubmed?term=Yamaguchi%20M%5BAuthor%5D&cauthor=true&cauthor_uid=10323487). Stimulatory effect on fracture healing of the femoral-diaphyseal tissues in rats. [Gen Pharmacol](http://www.ncbi.nlm.nih.gov/pubmed/?term=Stimulatory+effect+of+zinc+acexamate+administration+on+fracture+healing+of+the+femoral-diaphyseal+tissues+in+rats) 32(4):463-9, 1999.
21. [Jones L](http://www.ncbi.nlm.nih.gov/pubmed?term=Jones%20L%5BAuthor%5D&cauthor=true&cauthor_uid=20064837), [Thomsen JS](http://www.ncbi.nlm.nih.gov/pubmed?term=Thomsen%20JS%5BAuthor%5D&cauthor=true&cauthor_uid=20064837), [Barlach J](http://www.ncbi.nlm.nih.gov/pubmed?term=Barlach%20J%5BAuthor%5D&cauthor=true&cauthor_uid=20064837), [Mosekilde L](http://www.ncbi.nlm.nih.gov/pubmed?term=Mosekilde%20L%5BAuthor%5D&cauthor=true&cauthor_uid=20064837), [Melsen B](http://www.ncbi.nlm.nih.gov/pubmed?term=Melsen%20B%5BAuthor%5D&cauthor=true&cauthor_uid=20064837). No influence of alimentary zinc on the healing of calvarial defects filled with osteopromotive substances in rats. [Eur J Orthod](http://www.ncbi.nlm.nih.gov/pubmed/?term=No+influence+of+alimentary+zinc+on+the+healing+of+calvarial+defects+filled+with+osteopromotive+substances+in+rats)  32(2):124-30, 2010.

**Acknowledgements**

Our study was not supported by any Foundation

**Conflict of Interest**

The authors declare no conflict of interest

**Figure Legends**

**Fig 1:** Intragastrically supplement administration

**Fig 2; A:** Histopathological view of Group 1: Osteoblastic activity (Arrow), Haversian lamells (\*)(Hematoxylin and Eosin bar=50µm) **B:** Histopathological view of Group 2: Flat the osteoblast cells (Arrow), Myeloid tissues (\*) (Hematoxylin and Eosin bar=100µm) **C:** Histopathological view of Group 3: Bone loss in the defect area (Thin Arrow), Cubical osteoblast cells and Matrix formation (bold arrow) (Masson’s trichome Bar=100 µm) **D:** Histopathological view of Group 4: Trabecular matrix (Thin Arrow), Lipid cells (bold arrow) (Masson’s trichome Bar=100 µm)

**Fig 3:** Schematic summary of results. This diagram shows the median scores Osteoblastic Activity, Matrix Formation, Trabecular Bone Formation and Myeloid Tissue values of the Groups